Isolation of the New (-)-(3 R,4 S)-4-Hydroxymellein from the Fungus Septoria nodorum Berk

Michel Devys, Michel Barbier Institut de Chimie des Substances Naturelles, CNRS, Avenue de la Terrasse, 91198 Gif-sur-Yvette Cedex, France

Jean-François Bousquet, Albert Kollmann Station de Pathologie Végétale, CNRA, 78026 Versailles Cedex, France

Z. Naturforsch. **47 c**, 779–781 (1992); received April 6/July 7, 1992

Septoria nodorum (Phaeosphaeria nodorum), Fungus, Phytotoxins, Isocoumarins, (-)-(3 R,4 S)-4-Hydroxymellein

The new (-)-(3 R,4S)-4-hydroxymellein (6) is isolated from the fungus Septoria (Phaeosphaeria) nodorum Berk, together with the previously reported isomer 5. The results so far obtained in this series of metabolites are discussed in relationship with biosynthetic considerations.

Introduction

The fungus Septoria (Phaeosphaeria) nodorum Berk is a common parasite of wheat. It is responsible for a fungal disease which seriously affects crops (septoriosis). This fungus produces in cultures a series of highly potent phytotoxins which were previously investigated. Thus, we reported on the isolation of (-)-(3R)-mellein (ochracin) (4) [1]. (3 R)-O-methylmellein, (-)-(3 R,4 R)-4-hydroxymellein (5), (-)-(3R)-7-hydroxymellein, mycophenolic acid [2], septorine [3], N-methoxyseptorine [4], and N-methoxyseptorinol [5]. Undoubtedly, the observed phytotoxic activity of this fungus is due to the summation of these various metabolites, some of them being endowed of a particularly strong biological activity. Mellein was shown to produce an important reduction in CO2 net assimilation and an increase in stomatal resistance in seedling leaves of wheat [6-10].

In the present publication, the isolation of the new isomer (-)-(3R,4S) (6) from the culture medium of this fungus is reported. The results so far observed are discussed in relationship with the fact

Reprint requests to Dr. M. Barbier.

Verlag der Zeitschrift für Naturforschung, D-W-7400 Tübingen 0939–5075/92/0900–0779 \$01.30/0 that all the isocoumarins up to now isolated from Septoria nodorum are derived from (-)-mellein (4).

Results and Discussion

The culture medium of the fungus *Septoria* nodorum Berk was extracted by ethyl acetate according to a previously [11] reported method. The concentrated extract was submitted to HPLC fractionation (UV detection), leading to the known (-)-(3R,4R)-4-hydroxymellein (5) [2], plus the new compound (-)-(3S,4R)-4-hydroxymellein (6).

The structures of these substances 5 and 6 were determined on the basis of physicochemical data, by comparing with previously reported results (m.p. and $(\alpha)_D$ as represented in Fig. 1). In this series, the coupling constants of the protons at C-3 and C-4 are determinant for the attribution of the stereochemistry. As the absolute configurations established for (-)-mellein (1) and (+)-(3S,4S)-4-hydroxymellein (2) [12, 14], it was possible to fix the configurations of the products isolated from Septoria nodorum. The stereochemistry of the new product 6 was confirmed by irradiation at the signal of the methyl group at 1.55 ppm, resulting in two coalescent signals (2H) for the two vicinal protons at C-3 and C-4 (one peak). Hence, it is of course deduced that the coupling constant is not noticeable between these two protons. The dihedral angle determined from a molecular model gives a value of 70° which corresponds to a coupling constant of ca. 0.3 Hz [16]. By now, three of the four possible isomers of 4-hydroxymellein are known [2, 12, 14, 15]. To our knowledge, the (+)-isomer (3S, 4R) 3 still remains to be found and could exist in strains producing the corresponding (+)-mellein [14]. A diacetate of 6 was prepared which exhibited MS and ¹H NMR spectra in agreement with the proposed stereochemistry. In particular, the vicinal ³J coupling constant observed in the ¹H NMR of this diacetate is of approximately 1 Hz. It appears from the molecular models that two conformers are possible for substance 6, each requiring some energy for transformation. The dihedral angle 3-C-H 4-C-H is of ca. 70° when the methyl group is β-axial which leads to the observed coupling constant according to the Karplus equation [16]. But in the conformer having this methyl group β -equatorial, this angle is



780 Notes

Fig. 1. Absolute configurations of the two melleins (ochracins) and of the four possible 4-hydroxymelleins.

of 180° , a value which must give a higher J (between 6 and 15 Hz). Hence, two conclusions may be drawn: 1) the compound 6 isolated from Septoria nodorum has the 3-β-axial conformation, 2) the possibility that two conformers exist for each represented stereochemistry of 2, 3, 5, 6, can not be excluded. The observed differences in the $J_{3,4}$ -values between conformers of the cis-series 2 and 5 must be small (or negligible), whereas such differences should be much bigger in the trans-series 3 and 6.

The presence of the two isomers **5** and **6** in the culture medium of *Septoria nodorum* is in agreement with the demonstration that the biosynthesis of (+)-(3 S,4 S)-4-hydroxymellein (2) proceeds through oxidation of (+)-mellein (1) by molecular oxygen [14]. The previously found (-)-O-methylmellein and (-)-7-hydroxymellein, together present in *Septoria nodorum* with (-)-mellein [2] and the (-)-4-hydroxymelleins, demonstrate of course the same biogenetic family.

Experimental

Melting points have been determined with a Kofler apparatus under the microscope and are corrected. MS were carried out on an AEI MS 50 spectrometer, ¹H NMR on a Bruker 200 MHz apparatus (CDCl₃, δ ppm). Rotatory power was de-

termined on an electronic Perkin-Elmer polarimeter 241. TLC's on Schleicher-Schüll SiO₂ fluorescent films, 1 mm thickness for preparative purposes, UV observation with a Desaga lamp at 254 nm, or FeCl₃ spray for visualization.

The 4-hydroxymelleins 5 and 6 were extracted from the culture medium of Septoria nodorum according to the reported [11] method. The EtOAc concentration was submitted to HPLC (elution by ethanol-water, UV detection) and the two substances 5 and 6 recovered according to the absorption curve. SiO₂ TLC (CH₃Cl-EtOAc 5:1), 5 R₆ 0.60, colour reaction with FeCl₃: blue, 6 R_f 0.65, FeCl₃ pink, relative proportions 5:1. The physicochemical data of substance 5 are identical to the reported values [2] except for small differences in the m.p. and $(\alpha)_D$ due to more accurate technical facilities (such as electronic polarimeter). These values are reported on the Figure. 6: MS m/z (%), 194 M⁺ (100), 150 (70), 121 (80), 122 (80); ¹H NMR: 1.55 (d, 3H, CH₃), 4.50 (enlarged signal, 2H, H-3 and H-4), giving a singlet by irradiation of the methyl signal at 1.55; aromatic protons: 7.04 (d, 1 H, H-5, J = 8 Hz), 7.55 (dd, 1 H, H-6, J = 8 Hz), 6.95 (d, 1 H, H-7, J = 8 Hz), 11 (s, 1 H, 8-OH). The diacetate of 6 was prepared by treatment with acetic anhydride in the presence of anhydrous pyridine (10 mg 6, 6 drops of each reagent, 1 h at 37 °C, drying under in vacuo overnight at room temperaNotes

ture). SiO₂ TLC in pentane-ethyl acetate 1:1, R_f 0.70, MS m/z (%) 278 M⁺ (2), 236 M-42⁺ (100), 194 M-42-42⁺ (45), 176 M-42-60 (100), 150 (100), 149 (75), 43 CH₃CO⁺ (95). ¹H NMR (CDCl₃): 1.35 (d, 3H, CH₃), 2.13 and 2.40 (s, 3H each, CH₃COO), 4.80 (m, 1H, H-3), 5.83 (d, 1H, H-4, J = 1 Hz), 7.18 (d, 1H, H-5, J = 8 Hz), 7.36

(d, 1 H, H-7, J = 8 Hz), 7.66 (dd, 1 H, H-6, J = 8 Hz).

Acknowledgements

Thanks are due to Drs B. C. Das, C. Girard and J. P. Dupuis for the EI-MS determinations carried out at Gif-sur-Yvette.

- [1] M. Devys, J. F. Bousquet, M. Skajennikoff, and M. Barbier, Phytopathol. Z. **81**, 92 (1974).
- [2] M. Devys, J. F. Bousquet, A. Kollmann, and M. Barbier, Phytochemistry 19, 2221 (1980).
- [3] M. Devys, J. F. Bousquet, A. Kollmann, and M. Barbier, C.R. Acad. Sci. Paris 286, Ser. C, 457 (1978).
- [4] M. Devys, M. Barbier, A. Kollmann, and J. F. Bousquet, Tetrahedron Lett. 23, 4409 (1982).
- [5] M. Devys, M. Barbier, A. Kollmann, and J. F. Bousquet, Phytochemistry, in press.
- [6] J. F. Bousquet, H. Belhomme de Franqueville, A. Kollmann, and R. Fritz, Can. J. Bot. 58, 2575 (1980).
- [7] J. F. Bousquet, M. Skajennikoff, O. Bethenod, and P. Chartier, Ann. Phytopathol. 93, 503 (1977).
- [8] D. Laffray, J. F. Bousquet, O. Bethenod, and P. Louguet, Agronomie 2, 25 (1982).
- [9] O. Bethenod, J. F. Bousquet, D. Laffray, and P. Louguet, Agronomie 2, 99 (1982).

- [10] T. Touraud and J. F. Bousquet, Can. J. Bot. 57, 561 (1979).
- [11] J. F. Bousquet and M. Skajennikoff, Phytopathol. Z. 80, 355 (1974).
- [12] L. Camarda, L. Merlini, and G. Nasini, Phytochemistry 15, 537 (1976).
- [13] M. Sasaki, Y. Kaneko, K. Oshita, H. Takamatsu, Y. Asao, and T. Yokotsuka, Agr. Biol. Chem. 34, 1296 (1970).
- [14] C. Abell, A. C. Sutkowski, and J. Staunton, J. Chem. Soc. Chem. Commun. 1987, 586.
- [15] D. C. Aldridge, S. Galt, D. Giles, and W. B. Turner, J. Chem. Soc. (C) 1971, 1623.
- [16] L. M. Jackman, and S. Sterhell, Applications of NMR Spectroscopy in Organic Chemistry, p. 282, Pergamon Press, Oxford 1972.